THE CITRUS PSYLLA (DIAPHORINA CITRI, KUW.)
PSYLLIDAE: HOMOPTERA.

BY

MOHAMMAD AFZAL HUSAIN, M.Sc., M.A.,
Offg. Imperial Entomologist, Pusa.

AND

DINA NATH, L.A., B.Sc.,
Assistant, Entomological Section, Department of Agriculture, Punjab.

(Received for publication on 18th November 1920.)

INTRODUCTORY.

Of the Insect Pests of Citrus in India, Diaphorina citri, Kuw., commonly known as the "Citrus Psylla," * is of the greatest importance. It is present all over the country and has been so abundant in many localities as to call for special attention. In certain tracts in the Punjab it has been responsible for very serious losses. In one particular instance, a Citrus orchard which was normally leased for Rs. 2,000 per season yielded no more than Rs. 100 after an attack of this pest lasting for a couple of years. It is not an uncommon sight to see once valuable orchards reduced to unproductive plantations of dried skeletons of trees, primarily through the ravages of this insect (Pl. I. fig. 9).

This pest has been under observation for the last six years, both in the laboratory and under orchard conditions, and details of its structure, life-history, bionomics and control are presented in this paper. Crawford's and Ferris' have deplored the absence of detailed studies of the larval stages of Psyllids, and the importance of such accurate descriptions of the early stages as would permit of positive identification of insects cannot be overestimated in the case of pests. Valuable contri-

---

* In the Punjab this insect is commonly referred to as taila or chaipak. These terms are, however, very misleading, as they are applied indiscriminately to all those insects which secrete honey-dew. This sugary substance, falling on the leaves, gives them a shining appearance, as if smeared with oil, hence the term taila (taila: oil), and imparts to them a certain amount of stickiness, hence the term chaipak (chaipok: stickiness). All insects which belong to the families Fulgoridae, Jazzidae, Psyllidae, Aphididae Al-yroididae and Cocidae are spoken of as taila or chaipak.


butions to the study of the external morphology of the nymphal stages of a number of Psyllids have been made by Ferris¹, and the description of the immature stages of *D. citri*, with such details of the external features of the adult as may be useful, are included in this paper. Field observations and experiments which

![Image of attacked orchard: note dried branches.](image)

form Part II of this paper were carried out by the junior author under the guidance of the senior author. For the description of the various stages and the preparation of the paper for the press, the senior author is entirely responsible. This work was carried out during his tenure of office at the Agricultural Research Institute, Pusa.

Field investigations were carried out chiefly at Sargodha, Lyallpur and Gujranwala in the Punjab, and supplemented with observations from other parts of the Province, and confirmed by studies undertaken by the senior author at Pusa. The conclusions drawn will apply, with slight modifications, to other parts of India.

The photograph (Pl. II: fig. 5 d.) of the anal region of the nymph was taken by Mr. J. H. Walton, the Imperial Agricultural Bacteriologist, to whom our thanks are due for this help.

PLATE II.

*Diaphorina citri.*

1. Egg, side view.
2. First stage nymph, showing scolites of derm, dorsal view.
3a. First stage nymph, ventral view.
3. Second stage nymph, dorsal and ventral views.
4. Fourth stage nymph, dorsal and ventral views.
5. Fifth stage nymph, dorsal and ventral views.
5a. Margin of wing-pads, showing clavate setae.
5b. A lanceolate seta enlarged.
5c. Anus with circum-anal rings, highly magnified.
5d. Photograph of the anal region. (Anus is the lighter oval area, the inner pores appear as black dots, visible in places, and the outer ring of gland openings is distinct.)
PART I

SYSTEMATIC POSITION.

*D. citri*, Kuw., has been placed in the sub-family Psyllinae of the family Psyllidae (Chermidae) [Homoptera]. According to Crawford\(^1\) the principal and most distinctive characters of this sub-family are the concealed frons (Pl. III, figs. 1, 1a, fr), the presence of genal cones or lobate processes (g.c.), the presence of the apical claw-like spines on the posterior basal tarsus (not limited, however, to this group) (Pl. IV, figs. 3a, 3b, ta, cl), and the presence of the cubital petiole (M+C)\(^1\) (fig. 6). On the basis of the structure of the adult, *D. citri* certainly belongs to the sub-family Psyllinae, as it possesses very clearly all the above mentioned distinctive characters. A study of the immature stages of this species, however, reveals a different situation, as the nymphs differ markedly from all those that have, so far, been described from the sub-family Psyllinae, while, on the other hand, they bear a very strong resemblance to those of the sub-family Triozinae.

Crawford,\(^1\) for convenience, divides the sub-family Psyllinae into four tribes, and Ferris\(^2\) has studied the immature stages of the representatives of all these tribes:

- Tribe Pachypsyllini—*Pachypsylla venusta*, Osten Sacken.
- Tribe Euphyllyrini—*Euphyllyra arbuti*, Schwarz.
- Tribe Arytainini—*Psyllopsis fraxincola*, Foerster.
- Tribe Psyllini—*Psylla ulmi*, Linn., *P. buxi*, Linn.

The nymphs of the later stages of all these species agree in being of the typical Psylline type\(^2\)—the form in which the wing-pads are not produced cephalad at the humeral angle but project prominently from the contour of the body. The nymphs of *D. citri*, at any rate, the stages with wing-pads, do not resemble any of the above as the wing-pads are strongly produced cephalad at the humeral angle, and the nymphs therefore belong to the Triozine type\(^2\). They come very close to the nymphs of *Paratriozoa cockerelli*, Sule.,\(^2\) not only in the general form of the body and the cephalad prolongation of the wing-pads at the humeral angle, but also in the form and segmentation of the antenna—which are not more than 3-jointed even in the fifth stage nymphs, the structure of the legs and arrangement of the circum-anal pore-ring. Other nymphs of the sub-family Triozinae, which have so far been described, e.g., *Triozoa urticae*, Linn., *Phyllopecta dispyri*, Ashmead, and *Ph. tripunclata*, Fitch, are all modelled after the same pattern\(^2\) and agree with the nymphs of *D. citri*. Even *Ceropsylla sideroxylti*, Riley,\(^2\) which belongs to the Triozinae,

---

can easily be recognised as an exaggerated form of the same type—the Triozine type of Ferris.

In short, the nymphal affinities of D. citri suggest a closer relationship with the sub-family Triozinae rather than with the sub-family Psyllinae. A further study of the nymphs of the family Psyllidae is necessary to clear this point.

Franz Lowi (1878) erected the genus, to which this insect belongs, as Diaphora; but, as the name was preoccupied in Lepidoptera, he changed it to Diaphorina in 1879.

Crawford gives the following distinctive characteristics of the genus Diaphorina Low:

"Antennae very short, about as long as width of head or less, moderately thick. Genal cones thick, porrect (extending forward in same plane with vertex or nearly so). Thorax moderately narrow and slender, dorsal surface more or less granulate (gokornelt). Fore wings long, broadest near the apical end, basal half narrow; membrane more or less hyaline and usually maculated extensively with brown; pterostigma very narrow or wanting."

SYNONYMY.


DISTRIBUTION.

Diaphorina citri, Kuw., is widely distributed throughout the Orient. It is known from Formosa, the Philippine Islands, Moluccas (Ambon), Southern China (Macao), and occurs all over India. We have actual authenticated records from Pusa, Adra, Coimbatore and many localities in the Punjab. Specimens, in the collection of the Agricultural Research Institute at Pusa collected by T. Bainbrigge Fletcher at Poona on 8th September 1911, were first identified by Crawford as Euphalerus citri, but have since been found by him to belong to D. guttulata, Lethiery. Fletcher mentions E. citri from Cherat (N.-W. F. P.), but an examination of this single specimen shows that it is not D. citri, the wing pattern being quite different.

---

FOOD PLANTS.

*D. citri* has been found on all the cultivated varieties of *Citrus* and has been actually found attacking the following in the Punjab:

<table>
<thead>
<tr>
<th>Botanical name</th>
<th>English name</th>
<th>Local name</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Citrus aurantium</em></td>
<td>Orange</td>
<td>Sangtara, Malta</td>
</tr>
<tr>
<td><em>Citrus medica limonum</em></td>
<td>Lemon</td>
<td>Lima</td>
</tr>
<tr>
<td><em>Citrus medica limetta</em></td>
<td>Sweet lime</td>
<td>Mitha</td>
</tr>
<tr>
<td><em>Citrus medica asiaca</em></td>
<td>Sour lime</td>
<td>Khatta</td>
</tr>
<tr>
<td><em>Citrus medica medica</em></td>
<td>Citron</td>
<td>Galgal</td>
</tr>
<tr>
<td><em>Citrus doummana</em></td>
<td>Tumelo</td>
<td>Chakotra</td>
</tr>
</tbody>
</table>

*D. citri* has also been recorded from *Murraya koenigii* (Rutaceae) at Coimbatore, (S. India), and Pusa, and this may be considered as an alternative food plant. Crawford mentions *Euphorus citri*, Kew, as having been collected at Coimbatore by "T. V. R." on *Cordia* and by "C. N." on *Cardia* cordata (Boraginaceae), and Fletcher records it from *Cordia cordifolia* from the same locality. But the species found on *Cordia* spp. has since been identified by Crawford as *D. cardiae*. T. V. Ramakrishna Ayyar (1924) does not mention *Cordia cordata* among the plants on which *D. citri*, Kew, has been found, but on the other hand he records *C. cordata* as the food plant of *D. cardiae*. It may, therefore, be stated that so far *D. citri* has only been found on the plants of the order Rutaceae.

DESCRIPTION OF THE VARIOUS STAGES OF *D. CITRI*.

Egg.

(Pl. I, fig. 1, a, b, c; Pl. II, fig. 1.)

The egg may be described as almond-shaped, elongate, thicker at the basal end and tapering towards the distal end, where it is slightly curved and ends in a blunt point. From its rounded basal end arises a slender stalk-like process, which serves to anchor the egg, being thrust into the plant tissue. The length of the egg, without the stalk, is 0.3 mm., and the stalk measures 0.038 mm.; the greatest diameter of the egg is 0.13 mm. The colour of a freshly laid egg is pale yellow, but as development of the embryo proceeds, the colour gradually turns orange. A short time before hatching, two red spots—the eyes of the nymph—become visible. The egg shell is smooth and shining and no sculpturing of any description has been noticed. The arrangement of the egg membranes is the same as described for *Psylla mali* by Lees, and in general outline also there is a close

---

*It appears that Cordia has been mispelt as Cordine, the specific name of this insect may therefore be altered to Cordine.*

The distal end is
apt to impart to

Theating solution of
were obtained
permanent but
distinctly. To
horizontally
nymphs were
taffin. The
3; Pl. IV,
described by
cephalad
inge in the
green tinge,
dark, the
below:—
mm.
0.87
0.3
0.9
0.07

a distinct
sum of the
arrow strip
but ocelli
pinkish spots
quite large,
distinct
the process

Diaphorina citri (Citrus Psylla.)
PLATE III.

*Diaphorina citri* (Citrus Psylla).

4. Terminal portion of the antenna of the adult, ventral view.
5. Antenna of fifth stage nymph, dorsal view (structures on the ventral side shown separately and their position indicated by dots). *s.* sensorium.
6. Antenna, fourth stage nymph.
7. Antenna, third stage nymph.
8. Antenna, second stage nymph.
9. Antenna, first stage nymph.
10. Abdomen, male, terminal segments, lateral view (lettering as below).
15. Circuminal gland openings, highly magnified.
and the remaining portion not differentiated into segments, although some irregular faint lines are noticeable. From the proximal half of the undifferentiated portion of the antenna arise three lanceolate setae, the most proximal of which is the smallest. During life these setae are enclosed within cylindrical sheaths of wax similar to those described for the lanceolate setae of the abdominal region (see below). There are four sensoria on the ventral side of each antenna. In a cleared specimen the sensoria appear as hollow spheres. Besides these, there is a pit-like structure near the tip of the antenna, and a peculiar structure—which appears like a double ring—on the second joint. These two structures may also be sensory in function. A series of bristles arises from the ventral side of the antenna and its tip is armed with two unequal, thick, hollow bristles arising from separate protruberances. The *Rostrum*, as in the adult, is firmly attached to the selerite in the region of the prosternum, and is bent at a right angle between the coxae of the forelegs (in the diagrams it has been shown straightened out).

The *Thorax* (Pl. II, 5) is also distinctly marked and shows definite chitinized areas. The dorsum has three pairs of large selerites, a pair between the mesothoracic wing-pads and another between the metathoracic wing-pads; the third pair, which is the smallest, is, as it were, cut off from the first pair and is situated in front and near the middle line. Separating the larger selerites and the wing-pads are narrow strips of chitin.

The *Wing-pads* are very large and well developed and arise from the body by broad bases. The humeral angle is produced forwards reaching the level of the eye. Under very high magnification the surface as well as the margins of the wing-pads show small clavate hairs (figs. 5, 5a). Ventrally the thoracic region is soft except for a few thin strips of chitin. Two of these strips are transverse and are situated between the bases of the fore and middle legs, and the third runs longitudinally in the middle line between the middle and hind legs.

The *Legs* (Pl. II, fig. 5) are well developed and each consists of four segments, the trochanter not being differentiated. In all the legs the unjointed tarsus is separated from the tibia by a distinct joint. The end of the tarsus presents some interesting features. From its dorsal aspect just behind the tip arises a single, long, hollow seta, which may be described as golf-club-shaped,* as it has a long shaft and is bent and swollen at the end (Pl. IV, figs. 1, 1a, g. c. s.). The length of this seta is 0.07 mm. In the fifth stage nymph there is only a single seta on each tarsus. These setae evidently help in locomotion. If a nymph be observed under a binocular it will be noticed that the swollen end of the seta comes in contact with the surface on which the nymph is walking, and that the end of the seta remains attached to the surface a little after the claws and pulvilli have shifted forwards. These setae perform this function most probably by means of some sticky secretion. From under the end of the tarsus arises a fairly large fish-tail-shaped pulvillus (*p*).—

* We are indebted to Mr. Khan A. Rahman of the Entomological Section, Agricultural Research Institute, Pune, for suggesting this very expressive name for this seta.
the basal portion of which has a peg-like supporting structure. On the ventral side of the tarsus are two strongly chitinized median sclerites, the proximal of which has attached to it a long tendon which runs backward and can be traced up to the femur. The distal of these sclerites bears two strong bristles, the seats of origin of which appear as transparent circles. On the side of these median sclerites are the lateral sclerites which provide support for the paired claws.

The Abdomen (Pl. II, fig. 5) is well chitinized dorsally but is membranous ventrally. Four pairs of tergal sclerites can be distinguished and there is a chitinous apical plate composed of four or five pieces; faint grooves on this indicate segmentation. Between some of the anterior tergal sclerites there are thin chitinous strips which are intersegmental in position. Round the margin of the apical plate are a series of lanceolate setae. Their number varies from 53-60. These setae vary in size also but those towards the end are the longest and may be as much as 0.02 mm. in length. During life these setae are covered over with continuously growing, narrow, tubular sheath of waxy secretion. This secretion is produced by the glands situated round the bases of the setae, and the setae act as a central support for these small cylinders of wax. The ventral side of the abdomen is membranous except for a few thinly chitinized areas. The most marked of these is the plate bearing the anus and the circum-anal rings of pores and openings of glands. This plate is separated from the two lateral plates by folds, which run obliquely forward and also separate the small chitinous plates surrounding the spiracles from the sternites of the abdomen. The anal opening is well in front of the tip of the abdomen. As mentioned above, the anus is surrounded by a well-marked ring of minute pores (Pl. II, figs. 5c, 5d), and ornamentally arranged slits in a heavily chitinized area. Both these rings are interrupted anteriorly and posteriorly along the middle line. Two strong bristles are situated close to the posterior edge of the ring, on each side of the median line.

Fourth instar. (Pl. I, fig. 5; Pl. II, 4) Colouration and general form is the same as in the fifth instar; besides the size, the following differences may be noted:

There are only two lanceolate setae on the antenna, and only three sensoria; on the thorax only two pairs of dorsal sclerites are present, the smallest pair of the fifth stage not having separated as yet. The tarsi are not separated from the tibia.

Third instar. (Pl. I, fig. 4) Colouration slightly more orange; the general form of the body the same as for the above mentioned stages; the wing-pads are smaller, the anterior edge of the mesothoracic wing-pads just reaches the eyes; only the basal joint of the antenna is differentiated and there are only two sensoria and a single lanceolate seta.

Second instar. (Pl. I, fig. 3; Pl. II, fig. 3). Colouration and general form is the same as described above; the wing-pads are small triangular processes projecting from the body, the cephalad growth of the humeral angle is noticeable even at this stage; there is no lanceolate seta on the antenna and only one sensorium.
of which the second pair is cephalic. The cephalic
vein with the adjacent part of the hypopharynx is
shorter than the antennal segments.

First instar. (Pl. I, fig. 2, Pl. II, figs. 2, 2a). The nymph in this instar differs considerably from the later stages. Wing-pads cannot be made out, but the chitinous sclerites, which in later stages represent the wing-pads, are quite large. The arrangement of the abdominal sclerites is also very different. There are six pairs of tergites and the apical piece is probably made up of two pairs. Another peculiarity of this stage is that the dorsal surface of the sternum is a number of setae. Another feature in which the first-instar nymph differs from all the later stages is the presence of two golf-club-shaped tarsal setae on each of the middle and hind legs, and a single seta on each of the fore legs (Pl. IV, figs. 2, 2a). Further the pulvilli are not fish-
tail-shaped but are rounded at their free edges. The antenna is the same as described for the second instar nymph. The anus is nearer the apex of the abdomen than in the later stages. Besides the lanceolate setae around the abdomen there is one on each side of the thorax.

Comparative statement of the measurement of various parts of D. citri, during different nympha stages.

<table>
<thead>
<tr>
<th></th>
<th>I.</th>
<th>II.</th>
<th>III.</th>
<th>IV.</th>
<th>V.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Width of head in region of eyes</td>
<td>0.35</td>
<td>0.24</td>
<td>0.3</td>
<td>0.42</td>
<td>0.57</td>
</tr>
<tr>
<td>Length of antennae without terminal spines</td>
<td>0.06</td>
<td>0.08</td>
<td>0.12</td>
<td>0.18</td>
<td>0.3</td>
</tr>
<tr>
<td>Length of mesothoracic wing-pads</td>
<td>..</td>
<td>0.12</td>
<td>0.26</td>
<td>0.5</td>
<td>0.9</td>
</tr>
<tr>
<td>Length of metathoracic wing-pads</td>
<td>0.022</td>
<td>0.029</td>
<td>0.033</td>
<td>0.05</td>
<td>0.07</td>
</tr>
<tr>
<td>Length of body excluding lanceolate spines</td>
<td>0.26–0.35</td>
<td>0.13–0.56</td>
<td>0.47–0.72</td>
<td>0.76–1.2</td>
<td>1.6–1.7</td>
</tr>
<tr>
<td>Number of lanceolate spines on the apical plate</td>
<td>11–14</td>
<td>13–15</td>
<td>22–32</td>
<td>28–37</td>
<td>33–40</td>
</tr>
</tbody>
</table>

All measurements in mm.

Adult.

(Pl. I, III, IV).

The following is Crawford’s description of D. citri (Euphalerus citri):

- Length of body 2.4 mm.; length of fore wing 2.4 mm.; greatest width 1.0 mm.; width of vertex between eyes 3.5 mm.; with eyes 5.5 mm.; length of vertex to tip

---

of cones .32 mm. General colour brown, vertex slightly lighter; antennae black at tip; eyes darker, with black stripe from eye to tip of cone on side; forewings with a spotted macula of brown on the upper margin from a little below tip of clavus to tip of radius, and another on lower margin not quite merged into first, macula not continuous but composed of smaller scattered spots. Head narrower than thorax, descending; vertex flat, very finely pubescent, finely punctate, broader than long, with a lovea on each side in rear centre; anterior margin almost straight; facial cones broad at base, in same plane with vertex, convergent toward and rounded at apex, almost contiguous, finely pubescent. Antennal bases scarcely visible, as in other species; antennae short, not longer than head and pronotum combined, slender. Eyes large."

"Thorax very finely pubescent; pronotum rather long, sides parallel; pleurites typical for genus. Fore-wings subhyaline, rather thickened as in other species, maculate, attenuate at base, broadest subapically, rounded at apex, about two and a third times as long as broad; first cubital long; second marginal cell larger than first; radius long; pterostigma very narrow, quite long."

"Male—Genital segment rather conspicuous; anal valve elongate-pyriiform or flask-shaped, attenuate above; claspers long, slender, subacute at tip, simple."

"Female—Genital segment short, small, acute at tip; dorsal plate very slightly longer than ventral."

The above description was based on one female from Adra, Manbhum District (previously in Bengal now included in the province of Bihar and Orissa) collected
black at forewings ow tip of into first, narrower punctate, r margin convergent ventral bases head and pleurites species, two and a larger than uniform or ple."

PLATE IV.

Diaphorina citri (Citrus Psylla).

1. Tarsus, fifth stage nymph, ventral view. cl. claw, g.c.s. golf club seta, p. pulvillus; i, ii, iii, chitinous sclerites.
1a. Tarsus, fifth stage nymph, side view (lettering as above).
2. Tarsus, first stage nymph, ventral view (lettering as above).
2a. Tarsus, first stage nymph, side view (lettering as above).
3. Posterior leg of the adult. t.s. tibial spines.
3a. Lower part of tibia and tarsus of adult, ventral view. t.s.t. tarsal claws; t.s. tibial spines.
3b. Terminal portion of the basal joint of tarsus, highly magnified, to show the structure of pulvillus-like pad.
4. Middle leg of adult.
5. Fore leg of adult.
6. Fore wing. pt.a, pterostigma.

te very
District collected:
Diaphorina citri (Citrus Psylla).
by J. T. Jenkins (1909), and seventeen specimens of both sexes from the Philippines collected by George Compere. The description of the male was based on the Philippine specimens. Crawford has since examined more material from India.

The following notes are added to complete the above description and explain the diagrams.

The total length of the insect from the tip of genal cones to the apex of wings is 3·10—3·38 mm.; length of head from vertex to tip of genal cones is 0·28—0·34 mm.; width of vertex between eyes is 0·32—0·38 mm.; and width with eyes is 0·51—0·56 mm.

During life the insect is covered over with a whitish waxy secretion and appears as if dusted with white. The black stripe from eye to tip of genal cone is not distinctly visible in a living specimen but is quite clear in alcoholic material. Further, there is variation in the colour of this stripe; usually it is greyish, black only in very dark specimens. The colour of the abdomen also varies and appears to depend on the contents of the abdominal regions. It is usually greyish brown, but in some specimens it is distinctly bluish, while in gravid females, particularly during spring, it is distinctly orange.

The total length of the antenna is 0·58 mm., it is 10-jointed, the 3rd joint is the longest. At the distal ends of the 4th, 6th, 8th and 9th joints there are sensoria (Pl. III, figs. 2, 2a). The tip of the antenna is armed with two unequal hollow bristles.

The frons is concealed by the genal cones dorsally and is visible ventrally as a large plate (figs. 1, 1a).

The first two pairs of legs have fringes of whitish, delicate spines, at the end of tibia, the third pair have eight strong black spines. The tip of the basal joint of the hindmost tarsus has two claw-like spines and between these is a pad-like structure. (Pl. IV, figs. 3a, 3b).

The terminal segments of the abdomen are greatly modified for reproductive purposes. There exists considerable confusion regarding terminology used and difference of opinions concerning the segments which enter into the composition of these genital organs. The anus-bearing segment, which is known as the anal valve (Crawford)\(^1\) or supra-anal plate (Stough)\(^2\), is according to Wilmacl\(^3\) the tenth segment both in the male and female. Stough\(^2\) agrees with this interpretation and Singh Pruthi\(^4\) holds the same opinion in so far as male Psyllids are concerned. Crawford\(^1\) however, considers the anus-bearing segment to represent the eleventh segment.

---

segment in the male and female, the tenth segment being represented by the ovipositor of the female and the boat-shaped (sub-genital plates) segment of the male, in the hollow of which the aedeagus lies and to the posterior margin of which the parameres (Singh Pruthi) or forceps (Crawford) are attached. Witlaczil considers this to be the ninth segment, which is well developed ventrally and unrepresented dorsally. In the female, he considers the sub-genital plate (ventral genital valve) to be the tenth segment.

**Male.** (Pl. III, figs. 8, 8a, a. v.). In the male the tip of the abdomen is bent upwards. The anal valve is pyriform and hairy and the anus is carried at its end on a fleshy protuberance. This valve is carried at a right angle to the axis of the abdomen and forms the dorsal and anterior wall of the genital apparatus. The length of the anal valve is 0.36—0.4 mm.; Stough\(^1\) has given a full description of the male genitalia in *Pachypsylla c.-mamma*, and Singh Pruthi\(^2\) has also described these organs in a few Psyllids. The aedeagus (figs. 8, 9) consists of two parts, articulated in such a manner that the distal part is freely moveable, it is more or less straight and swollen slightly and bent at the tip. The cephalic face of the club is hollowed out to form a deep longitudinal groove. Arising from the front end of this groove is a tooth-like structure. The distal part of the aedeagus measures 0.2 mm. The proximal portion of the aedeagus is highly chitinized and strongly armed, it articulates with the base of the anal valve by broad plate-like expansions. The ejaculatory duct enters the aedeagus through the base of the proximal portion. This duct is chitinized and towards its anterior part it presents a spool-like structure (Pl. III, fig. 8). The middle portion of the spool is striated. The length of the spool is 0.060 mm.

Articulating with the base of the aedeagus is a loop of chitin and between the arms of the loop the proximal portion of the aedeagus lies. A median chitinous strut runs from the posterior end of the loop to the junction of the parameres.

The aedeagus and the loop and strut above-described lie in the hollow of the boat-shaped subgenital plate (or valve), which is well chitinized. The two flattened narrow parameres articulate with each other and with the posterior edge of the subgenital plate. Each of the parameres has a strong tooth at its extremity and is set with very long bristles, particularly on the median surface. The parameres are 0.3 mm. long.

**Female.** The end of the abdomen is not curved upward but is straight, coming to a point. The chitinized portion consists of two valves, the anal valve (supra-anal valve), which is dorsal, and the ventral subgenital valve. The dorsal valve is wedge-shaped and near its tip it is armed with short strong bristles (Pl. III, fig. 11,11b).

---


It is 0.7 mm. long. Near its base is a clear area and on this portion is situated the slit-like anus surrounded by an oval ring of a double row of the openings of wax glands. From these openings waxy secretion comes out. This elliptic ring is 0.16—0.2 mm. along its longest axis. The sub-genital valve (ventral valve) is boat-shaped and slightly shorter than the anal valve.

PART II

LIFE-HISTORY.

Many complete life cycles of this insect were studied by rearing adults on small citrus plants grown in pots and on tender shoots of plants in orchards. Attempts to rear nymphs on twigs cut off from the trees were not successful.

Copulation. Soon after emergence the adults copulate. As the genital segments of the male are turned upwards the male has to approach the female from the side and bend the tip of its abdomen down to the level of the aperture of the female genitalia; thus a male may be seen in that position holding the female with the legs of one side and supporting itself with the legs of the other side, the heads pointing in the same direction.

Oviposition. Soon after copulation, if tender shoots are available, the female starts laying eggs. The eggs are always laid on fresh growth, usually in the folds of half opened leaves, axils of leaves, pushed in between the buds and stem or petioles of leaves and axillary buds, and other similar situations. They are also laid scattered about on the upper or lower surfaces of the leaves and on tender stem. It may be stated that the females will not lay eggs on older tough leaves or stems even in the absence of tender shoots. The stalk of the egg is thrust into the tissue of the plant and the egg is firmly held in position. In some cases, however, particularly when too many eggs have been laid at the bottom of a folded or half opened leaf, a few of the eggs appear to be loose, simply glued on to the surface of the leaf. Such eggs come off when the shoot is placed in alcohol.

By virtue of their position the eggs are well protected from unfavourable climatic conditions, and as they are anchored into the plant tissue by means of their stalks they cannot be washed off by rain. Further, on hatching the nymphs obtain a rich supply of food.

Egg-laying records were obtained by confining, in sleeves placed on young plants, couples of freshly emerged adults. A single pair was placed in each sleeve. In captivity a female may lay over 800 eggs, and continue egg-laying for about two months. The statement below gives details about oviposition:—
THE CITRUS PSYLLA

**Daily egg laying records of *D. citri* during March, April and May (at Lyallpur).**
(The period of greatest activity.)

<table>
<thead>
<tr>
<th>Date</th>
<th>March 1923</th>
<th>April 1923</th>
<th>May 1918</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>A.</td>
<td>B.</td>
<td>C.</td>
</tr>
<tr>
<td>1</td>
<td>0</td>
<td>Em.</td>
<td>0</td>
</tr>
<tr>
<td>2</td>
<td>10</td>
<td>4</td>
<td>Em.</td>
</tr>
<tr>
<td>3</td>
<td>21</td>
<td>11</td>
<td>10</td>
</tr>
<tr>
<td>4</td>
<td>25</td>
<td>12</td>
<td>7</td>
</tr>
<tr>
<td>5</td>
<td>29</td>
<td>25</td>
<td>21</td>
</tr>
<tr>
<td>6</td>
<td>42</td>
<td>12</td>
<td>25</td>
</tr>
<tr>
<td>7</td>
<td>10</td>
<td>32</td>
<td>2</td>
</tr>
<tr>
<td>8</td>
<td>15</td>
<td>15</td>
<td>6</td>
</tr>
<tr>
<td>9</td>
<td>0</td>
<td>25</td>
<td>5</td>
</tr>
<tr>
<td>10</td>
<td>0</td>
<td>25</td>
<td>1</td>
</tr>
<tr>
<td>11</td>
<td>16</td>
<td>14</td>
<td>2</td>
</tr>
<tr>
<td>12</td>
<td>17</td>
<td>5</td>
<td>0</td>
</tr>
<tr>
<td>13</td>
<td>0</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>14</td>
<td>15</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>15</td>
<td>29</td>
<td>30</td>
<td>4</td>
</tr>
<tr>
<td>16</td>
<td>21</td>
<td>25</td>
<td>10</td>
</tr>
<tr>
<td>17</td>
<td>35</td>
<td>20</td>
<td>7</td>
</tr>
<tr>
<td>18</td>
<td>0</td>
<td>35</td>
<td>6</td>
</tr>
<tr>
<td>19</td>
<td>35</td>
<td>50</td>
<td>64</td>
</tr>
<tr>
<td>20</td>
<td>23</td>
<td>23</td>
<td>3</td>
</tr>
<tr>
<td>21</td>
<td>23</td>
<td>23</td>
<td>15</td>
</tr>
<tr>
<td>22</td>
<td>18</td>
<td>18</td>
<td>15</td>
</tr>
<tr>
<td>23</td>
<td>13</td>
<td>14</td>
<td>18</td>
</tr>
<tr>
<td>24</td>
<td>17</td>
<td>17</td>
<td>18</td>
</tr>
<tr>
<td>25</td>
<td>15</td>
<td>17</td>
<td>13</td>
</tr>
<tr>
<td>26</td>
<td>16</td>
<td>16</td>
<td>12</td>
</tr>
<tr>
<td>27</td>
<td>7</td>
<td>9</td>
<td>15</td>
</tr>
<tr>
<td>28</td>
<td>2</td>
<td>13</td>
<td>25</td>
</tr>
<tr>
<td>29</td>
<td>6</td>
<td>19</td>
<td>16</td>
</tr>
<tr>
<td>30</td>
<td>2</td>
<td>9</td>
<td>7</td>
</tr>
<tr>
<td>31</td>
<td>0</td>
<td>7</td>
<td></td>
</tr>
</tbody>
</table>

**Em.** = emerged.

**Ev.** = escaped.

**D.** = died.

*A* emerged on 28-h. but did not start oviposition till 1-iii. 23.

†A stopped egg-laying on 27-iv. 23, but lived till 5-vi. 23.

‡E stopped egg-laying on 27-iv. 23, but lived 6-vi. 23.

---

**Hatching.** The egg stage occupies from 4 to 6 days in summer and up to 22 days in winter. The nymph comes out of the egg shell through a slit that appears at the
narrower end of the egg. Lees\textsuperscript{1} has described a weak spot at the place where the egg-shell bursts. It takes about 20 minutes for the nymph to come out of the egg-shell completely. The empty egg-shell shrivels up into a scale-like structure and remains attached to the plant.

\textit{Nymphal instars.} There are five nymphal instars. In the younger stages the nymphs are mostly concealed, and are found at the tips of tender shoots. During the later stages they move down the shoot. They do not, however, leave the fresh shoot and are never found on last season’s growth. When disturbed, they move about in a slow, steady manner. They can withdraw their long mouth-parts and re-thrust them into the plant tissue quite readily.

Total duration in the nymphal instars varies from 11 to 25 days according to the season, and each instar occupies about 3 days in summer and 4 days in winter, except the first instar which occupies 11 or 12 days in winter.

\textit{Statement showing the duration of nymphal instars of D. citri during Summer and Winter.}

<table>
<thead>
<tr>
<th>First instar</th>
<th>28-iv</th>
<th>28-iv</th>
<th>28-iv</th>
<th>28-xii</th>
<th>24-xii</th>
</tr>
</thead>
<tbody>
<tr>
<td>Days</td>
<td>3</td>
<td>3</td>
<td>4</td>
<td>3</td>
<td>3</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Days</td>
<td>3</td>
<td>3</td>
<td>4</td>
<td>3</td>
<td>3</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Third instar</th>
<th>30-iv</th>
<th>4-v</th>
<th>4-v</th>
<th>4-v</th>
<th>4-v</th>
</tr>
</thead>
<tbody>
<tr>
<td>Days</td>
<td>3</td>
<td>3</td>
<td>2</td>
<td>3</td>
<td>4</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Fourth instar</th>
<th>30-iv</th>
<th>4-v</th>
<th>4-v</th>
<th>4-v</th>
<th>4-v</th>
</tr>
</thead>
<tbody>
<tr>
<td>Days</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>4</td>
<td>5</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Fifth instar</th>
<th>4-v</th>
<th>6-v</th>
<th>6-v</th>
<th>6-v</th>
<th>6-v</th>
</tr>
</thead>
<tbody>
<tr>
<td>Days</td>
<td>3</td>
<td>2</td>
<td>4</td>
<td>3</td>
<td>4</td>
</tr>
</tbody>
</table>

| Total duration | 14    | 13    | 15    | 26    | 28    |

\textit{Life-cycle.} From the statement below it will appear that the egg-stage may occupy 3 to 23 days, according to season, the nymphal stages 11 to 25 days, and the adult starts egg-laying soon after emergence. The total life-cycle occupies from 15 to 47 days. The adults live for a long time and have been known to live as long as 189 days.

SEASONAL HISTORY.

Eggs, nymphs and adults of *D. citri* are found on Citrus plants throughout the year, and no definite hibernation in any particular stage has been observed. During winter, which is very severe in most parts of the Punjab, the life-cycle is much prolonged (see above) and the insects are not very active except during the warmer parts of the day.

Adults have been known to live throughout the winter and lay eggs only in the spring when the new leaves have appeared. The Psylla becomes active about the end of February and its numbers increase very rapidly during March and April, when numerous eggs are laid and large colonies of nymphs in all stages of development are found on young shoots, flowers and new leaves. During May and June the insects are found mostly as adults, and in July—when the rains start and there is another outburst of fresh growth—these congregate on tender shoots and lay eggs, and nymphs hatch out in large numbers. The pest begins to disappear about the middle of October and during December and January it is present in very small numbers. This is due partly to the low temperature which retards development and partly to attacks of parasites which have been active all this time. (The facts mentioned above are true for Pusa also.)

The number of generations in the year. To find out the number of generations in the year observations were started on 1st July 1922 and continued till 29th June 1923. Eggs observed on an orange shoot were sleeved *in situ*. When adults emerged they were sleeved on to fresh shoots which had been previously
examined and found free from all stages of the pest. The following Table shows the numbers of generations which were actually reared:—

<table>
<thead>
<tr>
<th>Number of generations</th>
<th>Date of oviposition</th>
<th>Date of hatching</th>
<th>Date of emergence</th>
<th>Duration of life-cycle (days)</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1-vii-22</td>
<td>4-vii-22</td>
<td>18-vii-22</td>
<td>18</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>20-vii-22</td>
<td>23-vii-22</td>
<td>8-vii-22</td>
<td>19</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>10-vii-22</td>
<td>13-vii-22</td>
<td>30-vii-22</td>
<td>20</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>1-x-22</td>
<td>10-x-22</td>
<td>28-x-22</td>
<td>28</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>6-i-23</td>
<td>26-i-23</td>
<td>16-i-23</td>
<td>38</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>17-ii-23</td>
<td>24-ii-23</td>
<td>8-iv-23</td>
<td>22</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>16-iv-23</td>
<td>22-iv-23</td>
<td>3-v-23</td>
<td>17</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>23-v-23</td>
<td>27-v-23</td>
<td>7-vi-23</td>
<td>15</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>9-vi-23</td>
<td>14-vi-23</td>
<td>29-vi-23</td>
<td>20</td>
<td></td>
</tr>
</tbody>
</table>

Thus in these experiments nine generations were definitely traced during the year. It is, however, interesting to note that the adults which emerged at the end of August did not lay eggs for one month, i.e., until October, and those which emerged at the end of this month did not oviposit for over three months. As mentioned above, the insect does not undergo hibernation and in nature all the stages are met with throughout the year. Failure to lay eggs is, therefore, most probably attributable to the fact that during these periods young shoots were not available. If this interpretation is correct, then there is a possibility of at least two or three more generations during the year. As adults live for a very long time and continue egg-laying for a period which may extend to over two months there is considerable overlapping and no definite broods can be recognized.

HABITS AND BEHAVIOUR.

Habits of the nymphs. The young nymphs remain congregated on the young shoots close to the seat of oviposition. For a few days they continue to feed inside the unopened leaves, between the axillary bud and the stem or on the tender portion of the stem itself. When overcrowded, they crawl down the stem and gain sustenance from larger leaves and thicker parts of the tender stem. They are, however, confined to the fresh shoots. As a rule nymphs do not move much, but when disturbed or when migrating to a lower portion of the stem, they walk with a quick steady pace. The nymph sucks cell sap and exudes from its anus a thick sugary liquid which is covered over with a waxy secretion of the circumanal glands. This exudation appears as a thick translucent cord. When it has attained a certain size, the insect moves its abdomen from side to side in a characteristic fashion as if rubbing off this matter, and continues to do so till the wax-covered globule of honey-dew drops off. In cases of bad attack the ground under the trees appears white on...
account of the drops of this sugary excretion. When this matter falls on the leaves it gives them a shining appearance and stickiness. Later on fungus grows on this sugary material and the leaves appear to be covered over with a black sooty deposit.

Habits of the adults. The adults do not fly very far. They are usually found, in large numbers, sitting on the lower sides of the leaves with their heads almost touching the surface and the rest of the body raised up. The axis of the body thus forms an angle of 30° with the leaf. They do not possess the power of sustained flight and, unless disturbed, they do not move. Their movement consists of a leap in which the hind legs play an important part and the wings act as parachutes. When fresh leaves appear, the adults congregate on these, moving about quickly as if surveying the situation for oviposition. It has been noticed that if placed in a glass tube the adults move towards the window and congregate towards the top of the tube. Thus in captivity they are positively heliotropic and negatively geotropic. The migration of adults to fresh shoots, however, does not seem to be a response to either of these two stimuli. It may be repeated that adults will not lay eggs except on tender portion of the shoots.

SEASONAL HISTORY OF THE CITRUS TREES.

The Citrus trees are evergreen and continue to produce fresh leaves throughout the greater part of the year. The seasons for flushing, however, in the Punjab and at Fusa are the spring (end of February to April) and the rainy season (July and August), when fresh shoots appear and leaves are produced in great abundance. In most varieties the floral buds begin to appear during the last week of February and flowers continue to open till about the end of March. The fruit sets in March. In the case of oranges and malant, the fruit remains on the tree for the rest of the year ripening in November and December and developing its finest flavour in January and February of the following year. The acid varieties ripen earlier.

The exact dates of flushing, flowering and fruiting vary with the locality, season, and variety, but the above may be taken as the typical seasonal history of the host. In some varieties not only the leaves but flowers and fruits continue to appear and fruits of all sizes are present on the plant throughout the year.

DAMAGE.

D. citri neither produces galls nor causes any malformation of the plant tissues. The only external sign of injury is defoliation and death of the shoots attacked and the drying up of the branches. On badly infested trees the continual feeding of myriads of insects is, in itself, a very great drain on the food-supply of the plant, but most probably some poison is also injected into the tissues of the host and this produces the more serious results. This is evident from the fact that the fruit of
the infested tree is dry and insipid to taste, and branches other than those actually attacked also dry up. Besides, young shoots are killed, old leaves fall off and in course of time an attacked tree is demided of leaves altogether. The period of the greatest activity of the insects and most rapid increase in their numbers corresponds with that of the sprouting of new shoots and appearance of new leaves. The greatest damage is therefore done at this stage. In the Punjab it is during March and April that the attack is most severe, and every fresh crop of leaves and buds is killed. During the first year of attack the damage is not very marked and the trees do not appear much affected. The yield, however, falls, and some top branches dry up. During the second year of attack, if the pest has been allowed to develop unchecked, all the new shoots are destroyed, and most of the branches are left without leaves and the tree begins to dry up. Very little fruit is borne and that too is of small size, insipid and dry. During the third season, there is neither leaf nor fruit on the trees and, if no remedial measures are employed, the trees begin to die. At Sargodha where the pest had been allowed to multiply unchecked for several years, it was not an uncommon sight to see large trees reduced to dry twigs without a leaf.

The citrus growers, being ignorant of the potentialities for damage possessed by these small insects, attribute the failure of crop to other causes and the real culprits remain undetected for a long time and are often only noticed when they have established a firm foothold. The extent of damage which D. citri is capable of doing may be judged from the following two cases which were carefully studied and for which figures are available:

<table>
<thead>
<tr>
<th>Locality</th>
<th>Year</th>
<th>Income</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Sargodha</td>
<td>1915</td>
<td>400</td>
<td>Average income per year before the attack.</td>
</tr>
<tr>
<td></td>
<td>1916</td>
<td>250</td>
<td>Serious attack during this year.</td>
</tr>
<tr>
<td></td>
<td>1917</td>
<td>23</td>
<td>Severe attack.</td>
</tr>
<tr>
<td></td>
<td>1918</td>
<td>400</td>
<td>Orchard sprayed in September 1917 and March 1918.</td>
</tr>
<tr>
<td></td>
<td>1919</td>
<td>500</td>
<td>No attack.</td>
</tr>
<tr>
<td></td>
<td>1920</td>
<td>700</td>
<td>No attack.</td>
</tr>
<tr>
<td></td>
<td>1915</td>
<td>1,200</td>
<td>Normal income before the attack.</td>
</tr>
<tr>
<td></td>
<td>1916</td>
<td>500</td>
<td>Serious attack.</td>
</tr>
<tr>
<td></td>
<td>1917</td>
<td>120</td>
<td>Very little fruit; attack very severe.</td>
</tr>
<tr>
<td></td>
<td>1918</td>
<td>700</td>
<td>Orchard sprayed once during March 1918.</td>
</tr>
<tr>
<td></td>
<td>1919</td>
<td>500</td>
<td>Orchard not sprayed.</td>
</tr>
<tr>
<td></td>
<td>1920</td>
<td>1,300</td>
<td>Orchard thoroughly sprayed early in the season.</td>
</tr>
</tbody>
</table>

PREDATORS AND PARASITES.

So far we have not obtained any parasites from the eggs of D. citri and no predators have been noticed. The adult also seems to be fairly free from enemies. The
nymphal stages, however, are subject to the attacks of a number of predators and parasites, which play a very important part in keeping this pest in check.

**Predators.** Of the more common predators on *D. citri* nymphs the following may be mentioned.:

*Coccinellidae*:

*Coccinella septempunctata*, Linn., is a very common species. It usually feeds on Aphids. Both the larvae and adults have been noticed to feed on the nymphs of this Psylla. A complete life-cycle occupies about 17 days. These beetles, therefore, multiply rapidly and are capable of doing enormous good.

*C. repanda* has similar habits. It, however, does not occur in such large numbers as the above mentioned species.

*Chilomenes sermaculata* occurs in fairly large numbers and does useful work.

*Chilocorus nigrita* also occurs in very large numbers feeding on the nymphs of Psylla.

*Brusius naturalis* is a very small beetle. It is not very common and has been noticed in only a few orchards.

*Syrphidae*: A Syrphid fly was also found in large numbers, the maggots of which are observed feeding on the larger nymphs. In March and April, when its food supply is abundant, the entire life-cycle occupies 12 days.

*Chrysoptidae*: The larvae of *Chrysopa* sp. were also present in large numbers feeding on the nymphs.

Besides the above, certain Spiders and Mites are also found feeding on adults of Psylla.

**Parasites.** The parasites of the nymphal stages, however, play a more important part than the predators. So far nine different Chalcidoid parasites have been reared from the nymphs of *D. citri*. The most important of these is *Tetrastichus radiatus*, Waterston1. The percentage of parasitization is often very high and as many as 95 per cent. of the nymphs may be attacked. From the observations made by us it seems likely that in all those localities where *D. citri*, although present, does not increase in numbers sufficiently to cause any serious damage, the credit of keeping the pest in control is mainly due to this parasite. Unfortunately a number of hyperparasites have also been noticed.2

**CONTROL MEASURES.**

As mentioned above, the predators and parasites play a very important part in keeping this pest in check, and every effort should be made to increase the number of these beneficial insects.

For immediate relief, however, spraying is recommended. Care should be taken that this measure is not employed at a time when the percentage of parasitization is very high. Spraying has been tried extensively and has proved thoroughly
decocitive work we also give expensive manners.
A set of water 4 chahs dissolve water for
Tobacco does not

Ros spraying

Ros


---

effective. Any good contact poison will serve the purpose. We recommend tobacco decoction and rosin compound, as these can be prepared on the spot, are easy to work with and the ingredients are available everywhere. Crude oil emulsion has also given very good results, but, taking into consideration the freight, it becomes expensive. During our experiments tobacco decoction was prepared in the following manner:

A *seer* (2 lb.) of tobacco refuse (leaves, stems, dust, etc.) was soaked in 10 seers of water for 24 hours, after which it was strained, and to the liquid was added about 4 *chataks* (½ lb.) of cheap soap cut into small pieces, the solution being stirred to dissolve the soap thoroughly. This stock solution was diluted with 8 parts of water for spraying purposes.

Tobacco decoction is more useful for spring spraying against the nymphs. It does not injure the plant at all.

Rosin compound is particularly efficacious against the adults during winter spraying. It is prepared according to the following formula:

- **Rosin**, 2 seers.
- **Soda**, 1 seer.
- **Sollignum or Sanitary fluid**, 1 seer.
- **Water**, 4 gallons.

Rosin and soda are placed in a vessel and sufficient water to cover is added, and the whole is heated over a slow fire till it boils, and the boiling is continued till rosin and soda are completely dissolved. During the process water is added slowly all the time, until 4 gallons have been added. The compound is tested from time to time to see if it mixes well with water. Before it is fully cooked a drop of it when put in water forms a milky emulsion; when, however, it is ready it mixes without producing milkiness. This stock solution is diluted with 6 parts of water. Sollignum is added to the solution and well mixed.

During the spraying operations in the Punjab a Four Oaks Lime washing machine was used, and on an average a gallon of the spraying mixture was quite enough for one tree.

**Spraying schedule.** It is very important that spraying should be done at the right time.

A thorough spraying with rosin compound during the winter will kill most of the adults. The trees must be very carefully sprayed and a good pressure maintained, and the underside of the leaves where insects congregate must receive special attention. It may be necessary to repeat the operation. If Winter spraying has been done satisfactorily there will be no need to undertake Spring spraying, but in case a few adults have survived or have migrated from other orchards, they must be dealt with. This should be undertaken when most of the nymphs have hatched out and the adults of the new brood have not yet emerged. In the Punjab, March is the best time for this. Tobacco decoction has given very good results. As the nymphs are mostly concealed in the fresh shoots, spraying should be directed towards
these parts. A very fine mist should be produced so that the insecticide reaches everywhere. The eggs are more difficult to kill, but if the attack is serious, a third spraying may be done after about a week or so, to kill any nymph that may have hatched.

During 1925-26 about 5,000 trees were sprayed and the efficacy was from 74—95 per cent., rosin compound giving the best results. (Text fig. 3)

![Image of a branch after spraying showing dead hoppers.](image)

**PREVENTIVE MEASURES.**

The plants which receive proper cultivation are able to withstand the attacks of pests much better than sickly plants, and it is advisable that attention be paid to cultural methods in pest control.
Citrus hedges are commonly grown in most parts of the Punjab. A well-trimmed hedge is always producing fresh leaves which provide food to the nymphs and on which the adults oviposit. These hedges, therefore, serve as reservoirs for the pests, and in the vicinity of citrus orchards such hedges should be discouraged.

The nurserymen who sell Citrus plants to the orchard owners have no idea of the importance of selling pest-free stock, and it is not uncommon to find seedlings covered with nymphs of Psylla being offered for sale and bought. Steps should be taken to legislate against the sale of plants attacked by Insect Pests.

**SUMMARY.**

*Diaphorina citri*, Kuw., is a serious pest of Citrus plants in many parts of India. The pest attacks young tender shoots and kills them. In course of two or three seasons, the tree is completely defoliated and begins to dry up. During severe attacks the fruit does not set or drops off, and what is left is dry and insipid.

Eggs are deposited on the young shoots, usually in clusters within the folds of unopened leaves, between the petioles of leaves and buds, and buds and stem. The nymphs that hatch out remain congregated on these tender shoots and kill them. Eggs are not laid on old leaves and the nymphs are never found except on the fresh growth. There are five nymphal instars and the life cycle takes from 15 to 20 days in summer. There are about nine generations in the year. The female may lay up to 800 eggs and therefore the increase in numbers is very rapid.

Several different predators and parasites help to keep this pest in check, and of particular importance is *Tetranychus radiatus*, a Chalcidoid parasite.

Spraying is an effective method of control; rosin compound and tobacco decoction have given good results.

Attention should be paid to proper cultivation and Citrus hedges should not be grown around Citrus orchards, because these hedges act as reservoirs for fresh infestations of Citrus Psylla.