

# Integrating quantitative and molecular genetics in host-range determination

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# Mission of Foreign Disease - Weed Science Research Unit

- Find and develop **foreign** plant pathogens for classical biological control of introduced invasive weeds in the U.S.

# Steps in Developing Plant Pathogens for Classical Biological Control of Weeds

- Find diseases of introduced weeds in the native (foreign) ranges of the weeds
- Isolate the pathogens
- Test the pathogens for damage
- Test the pathogens for their host range (safety)

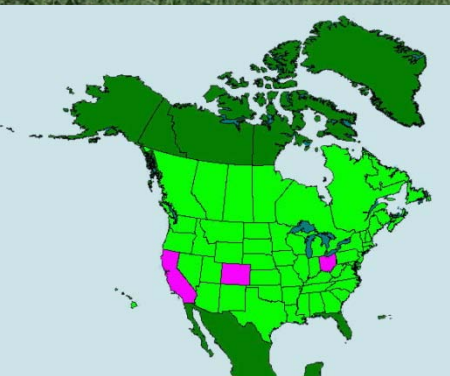
## Target:

- ▶ Russian thistle (*Salsola tragus*)

## Pathogens:

- ▶ *Colletotrichum gloeosporioides* f. sp. *salsolae* (CGS)  
Facultative parasitic fungus  
Hungary (other isolates from Greece and Russia)
- ▶ *Uromyces salsolae*  
Obligate parasitic rust fungus  
Russia

# Tumbleweed/Russian Thistle



*Colletotrichum gloeosporioides* f. sp. *salsolae* (CGS)



*Uromyces salsolae*



# Problem

**How to best evaluate disease reaction of non-target plant species?**

- Avoid discarding potentially beneficial organisms**
- Ensure pathogen safety**



# Partial Disease Severity Results (CGS)

Table. Plants in the Chenopodiaceae

Genus	Species	No. Plants		Average Disease Rating
		Inoc	Pos	
Grayia	spinosa	10	0	0
Kochia	scoparia	15	0	0
Nitrophila	occidentalis	7	7	1.8
Salicornia	bigelovii	1	1	4.0
Salicornia	maritima	5	5	3.6
Salicornia	virginica	5	0	0
Salsola	kali	11	9	2.4
Salsola	orientalis	13	2	0.4
Salsola	paulsenii	5	5	4.0
Salsola	tragus	261	252	2.9
Salsola	australis	65	55	1.1
Suaeda	californica	1	0	0
Suaeda	taxifolia	4	4	2.2

# NEED

- Environmentally-independent measure of disease severity
- Disease severity for species (vs. individual plants)
- An objective indicator of susceptible & non-susceptible species
- A means to evaluate rare and difficult or impossible to grow species
- A means to objectively compare disease on target vs. non-targets

# Partial Solution (next best)

- **Use ranks of disease severity ratings**
- **Use logit values of disease incidence**
- **Analyze using weighted mixed model ANOVA**
- **Produce least squares mean estimates**

# Full Solution

Combine disease evaluation data with another statistical approach

- Incorporate all available knowledge
- Incorporate genetic relationships among species with variances and disease evaluation data
- Get broad-based predictions for species

# Mixed Model Equations (MME) and Best Linear Unbiased Predictors (BLUPs)

- Long-established quantitative genetics approach
- Standard in generation of breeding values (BLUPs) for dairy cattle worldwide.
  - C. R. Henderson – 30+ years of publications from 1949
- Also for quarter horses, swine, trees, crop plants,.....
- Common objective:
  - Predict breeding values (genetic merit of potential parents) -BLUPs

# BLUPs

**Best** – minimum mean squared error

**Linear** – linear function of the data

**Unbiased** – average value of the estimate equals average value of quantity being estimated; no estimable function bias

**Predictor** – realized value of a random variable

BLUPs are also parametric Bayes estimates

# Value of the MME & BLUPs in Host Range Determination

- Predict susceptibility of plant species relative to that of the target species
- Use all available information
  - disease ratings or incidence
  - variance/covariance structure
  - genetic relationships among species
- Predict disease reactions for species that cannot be tested
- Determine the complete host-range of a pathogen among both tested and not tested species
- Determine relevant lists of non-target species to test

# How?

- Incorporate genetic relationships
- Incorporate genetic variances
- Incorporate performance (disease evaluation) data



# How?

Incorporate genetic relationships from:

- Pedigrees and coefficients of co-ancestry

OR

- DNA sequences and genetic distance matrices

# How?

Incorporate genetic variances from

- Heritability estimates

OR

- Estimate variance from data

# How?

**Incorporate performance data  
(disease/damage evaluation data)**

# How?

- Integrate data into Mixed Model Equations
- Generate Best Linear Unbiased Predictors (BLUPs) for species

## How?

- Generate DNA sequences, ITS (and/or other), for plant species
- Generate distance matrix among species based on these sequences
- Integrate these genetic distances with performance data
- Run the MME to generate BLUPs

# Model

## (in matrix notation)

- $y = XW + ZU + E$ 
  - $y = n \times 1$  vector of ranks of disease ratings
  - $X = n \times 1$  design vector of “1”s for the fixed intercept only
  - $W = 1 \times 1$  unknown vector for the fixed effect parameters, in this case only the intercept
  - $Z = n \times j$  design matrix for the random effects, in this case species
  - $U = j \times 1$  unknown vector of the random effects parameters
  - $E = n \times 1$  vector of residuals (errors)
- variance of  $y = S = ZGZ' + R$ 
  - $G = j \times j$  matrix of variances and covariances from distance matrix of DNA sequences and variance among species
  - $R = n \times n$  error matrix with known error variance as estimated from data
- $\hat{U} = GZ' \hat{S}^{-1} (y - X\hat{W}) = \underline{\text{BLUPs}}$

# Partial G matrix

relationship matrix=(1-each element of distance matrix)

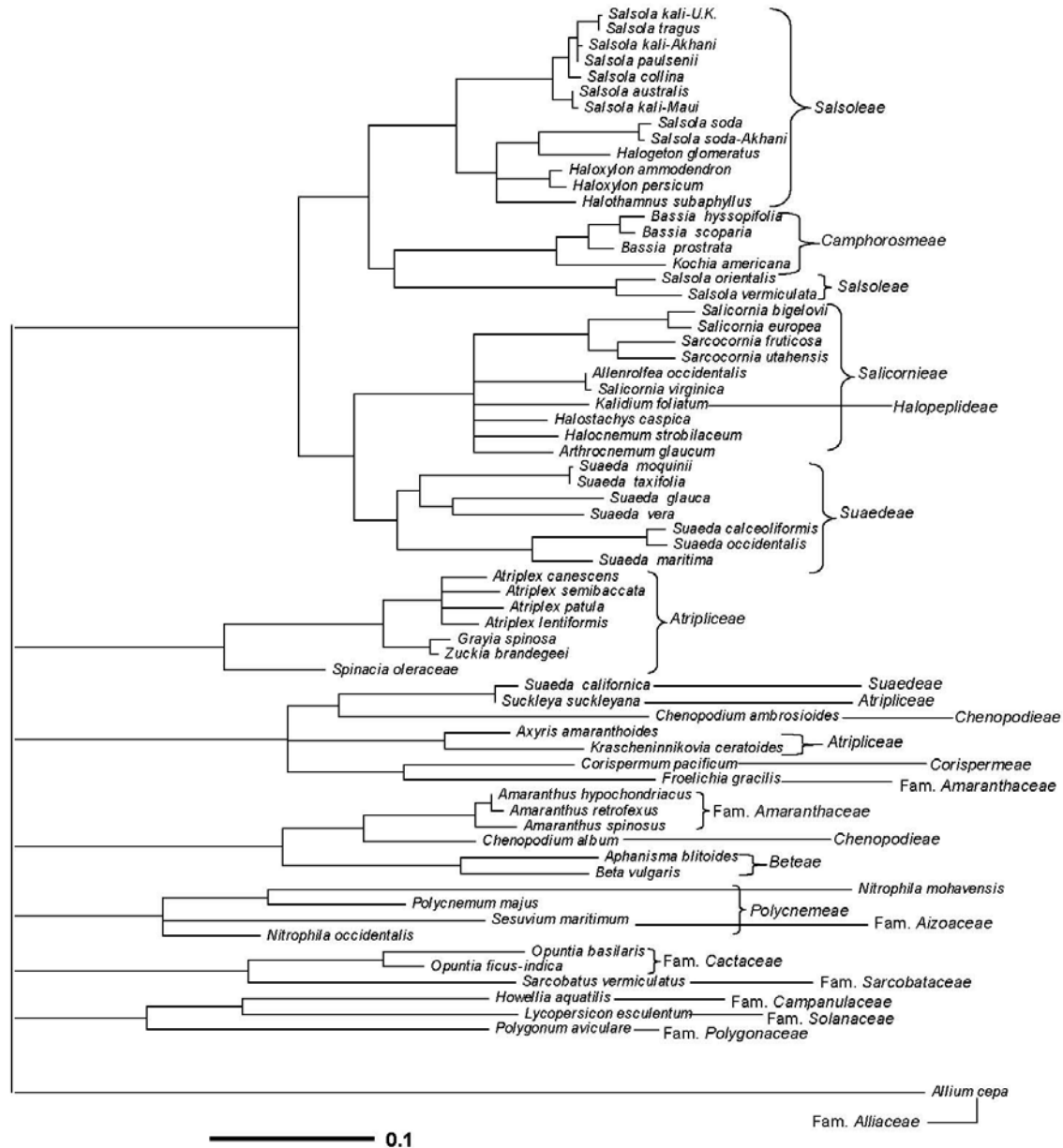
G= Variance among species × each element of relationship matrix

## The Mixed Procedure

### Estimated G Matrix

Row	Effect	taxon	Col1	Col2	Col3	Col4	Col5	Col6
1	taxon	Allenrolloccident	2324.43	2173.27	732.64	1617.11	1617.11	1626.61
2	taxon	Allenrolvaginata	2173.27	2324.43	797.44	1764.68	1764.68	1777.79
3	taxon	Allium cepa	732.64	797.44	2324.43	650.33	643.54	656.05
4	taxon	Amaranthcaudatus	1617.11	1764.68	650.33	2324.43	2324.43	2309.55
5	taxon	Amaranthhypochon	1617.11	1764.68	643.54	2324.43	2324.43	2309.55
6	taxon	Amaranthretrofle	1626.61	1777.79	656.05	2309.55	2309.55	2324.43
7	taxon	Amaranthspinosus	1606.60	1757.78	649.26	2252.49	2252.49	2234.94
8	taxon	Aphanismblitoide	1653.06	1845.36	628.13	1729.70	1725.17	1743.58
9	taxon	Atriplexcanescen	1773.08	1850.73	753.39	1688.16	1688.16	1714.27
10	taxon	Atriplexlentifor	1763.55	1832.70	739.17	1702.23	1702.23	1727.96
11	taxon	Atriplexpatula	1776.28	1802.67	732.08	1743.21	1743.21	1752.92
12	taxon	Atriplexsemibacc	1746.04	1789.23	755.56	1674.71	1674.71	1701.34
13	taxon	Bassia hyssopif	1738.95	1811.29	681.80	1519.57	1519.57	1534.22
14	taxon	Betavulgaris	1678.15	1873.30	770.69	1756.55	1751.99	1765.33
15	taxon	Bougainvsp	1448.21	1487.43	777.71	1299.52	1299.52	1317.56
16	taxon	Brassicaoleracea	907.18	1274.46	490.90	833.56	825.87	855.44
17	taxon	Calendulofficina	1097.90	1227.04	538.18	862.29	855.95	863.39
18	taxon	Carduus acanthoi	1094.11	1172.19	542.17	1011.59	1005.48	995.09
19	taxon	Carduus pyonocep	1175.63	1269.35	511.21	1045.76	1039.69	1045.04
20	taxon	Carthamutinctori	1124.93	1172.05	473.60	1003.39	997.32	992.69
21	taxon	Centaurecyanus	1192.36	1340.50	594.01	1052.29	1046.16	1048.57
22	taxon	Centaurediffusa	1205.73	1332.94	564.33	1067.38	1061.29	1063.75
23	taxon	Centauresolstiti	1176.07	1290.59	555.17	1046.30	1040.16	1042.58
24	taxon	Chenopodaalbum	1704.32	1783.93	879.26	2029.81	2029.81	2040.97
25	taxon	Chenopodambrosio	1520.15	1615.04	575.74	1435.99	1435.99	1469.92
26	taxon	Corispermipacificu	1711.90	1811.80	733.29	1609.67	1609.67	1619.01
27	taxon	Crupina	1112.26	1225.30	563.60	986.84	980.61	982.89
28	taxon	Cupressuabramsia	656.58	1090.99	131.31	573.32	579.02	573.09
29	taxon	Cupressugovgov	776.20	1307.58	361.01	645.84	651.49	645.10
30	taxon	Cynara scolymus	1199.24	1266.44	486.81	953.18	947.04	943.28
31	taxon	Daucus carota	1298.19	1372.51	750.74	1066.08	1066.08	1068.98
32	taxon	Endolepicovillei	1106.36	1185.95	547.26	949.13	942.91	978.72
33	taxon	Froelichgracilis	1558.34	1688.56	835.68	1494.70	1494.70	1478.55
34	taxon	Gossypubarbaden	1050.18	1252.47	374.77	883.19	876.89	885.63
35	taxon	Halocnemstrobila	2098.08	2016.84	687.85	1641.30	1642.86	1652.23
36	taxon	Halostaccaspica	2108.77	2101.45	941.95	1650.30	1650.30	1661.32
37	taxon	Halothamsubaphyl	1785.12	1844.81	861.18	1565.67	1565.67	1569.76
38	taxon	Haloxyluammodend	1611.25	1510.97	685.80	1359.91	1359.91	1363.72
39	taxon	Haloxylopersicum	1794.67	1891.44	832.73	1611.78	1611.78	1615.92
40	taxon	Howelliaaquatili	1300.45	1392.66	691.56	1264.47	1264.47	1280.48
41	taxon	Kalidiumfoliatum	2039.20	2021.77	647.47	1653.06	1653.06	1663.92
42	taxon	Kochia american	1716.50	1789.88	706.86	1487.36	1487.36	1487.70
43	taxon	Lycopersesculent	1266.23	1315.56	708.02	1208.47	1202.50	1204.52
44	taxon	Malaco fasci	1089.53	1249.71	375.88	989.21	983.21	991.83
45	taxon	Mirabilimultiflo	1341.54	1364.42	686.08	1231.53	1231.53	1224.95
46	taxon	Opuntia ficus	1538.33	1616.04	713.39	1405.09	1399.28	1410.7

# Phylogram from ITS sequence data





# Power

- Beta ( $\beta$ ) = probability of committing a Type II error and not rejecting a false null hypothesis
- Declaring no significant difference from zero when a difference exists
- Declaring a species not susceptible when it probably is
- Power =  $1 - \beta$ , probability of correctly rejecting a false null hypothesis
- Power values  $\geq 0.80$  are generally regarded as significant

# Least squares means

Genus species	CGS				<i>Uromyces salsolae</i>			
	Least squares means estimates	Standard error of estimate	Pr> t	Power (1-β)	Least squares means estimates	Standard error of estimate	Pr> t	Power (1-β)
<i>Salsola kali</i> -U.K.	285.40	41.47	<0.0001	0.167	132.10	10.89	<0.0001	0.082
<i>Salsola tragus</i>	277.82	37.46	<0.0001	0.202	141.59	3.71	<0.0001	0.132
<i>Salsola collina</i>	285.98	35.00	<0.0001	0.235	62.05	0.54	NS	0.781
<i>Salsola paulsenii</i>	296.40	34.19	<0.0001	0.248	120.34	44.09	NS	<0.050
<i>Salsola kali</i> -Akhani	NT	NT	-----	-----	NT	NT	-----	-----
<i>Salicornia bigelovii</i>	247.06	17.30	<0.0001	0.999	61.21	2.22	NS	0.321
<i>Salsola australis</i>	111.24	12.72	NS	0.996	60.20	3.38	NS	0.200
<i>Salsola kali</i> -Maui	111.08	18.00	NS	0.884	36.67	52.20	NS	<0.050
<i>Salicornia europaea</i>	144.18	16.09	NS	0.922	60.03	4.76	NS	0.148
<i>Sarcocornia fruticosa</i>	NT	NT	-----	-----	NT	NT	-----	-----
<i>Bassia hyssopifolia</i>	239.08	29.50	<0.0001	0.355	52.64	39.61	NS	<0.050
<i>Bassia scoparia</i>	110.60	10.35	NS	>0.999	NT	NT	-----	-----
<i>Nitrophila occidentalis</i>	254.37	81.79	NS	<0.100	60.20	3.38	NS	0.200
<i>Halothamnus subaphyllus</i>	NT	NT	-----	-----	NT	NT	-----	-----

59 species evaluated

6 susceptible (non-zero) species

CV= 3.31 to 407.87%

46 species evaluated

3 susceptible (non-zero) species

CV= 2.6 to 142.4%

# BLUPs

Genus species	CGS				<i>Uromyces salsolae</i>			
	BLUP	Standard error of prediction	Pr> t	Power (1-β)	BLUP	Standard error of prediction	Pr> t	Power (1-β)
<i>Salsola kali</i> -U.K.	247.92	10.23	<0.0001	>0.999	102.94	5.30	0.006	>0.999
<i>Salsola tragus</i>	246.73	9.84	<0.0001	>0.999	101.97	5.10	0.007	>0.999
<i>Salsola collina</i>	235.40	10.70	<0.0001	>0.999	95.61	5.33	0.019	0.995
<i>Salsola paulsenii</i>	225.51	7.59	<0.0001	>0.999	96.90	4.65	0.023	>0.999
<i>Salsola kali</i> -Akhani	224.75	7.93	<0.0001	>0.999	97.02	4.85	0.023	>0.999
<i>Salicornia bigelovii</i>	213.39	14.95	<0.0001	>0.999	54.47	7.93	NS	>0.999
<i>Salsola australis</i>	208.14	10.29	<0.0001	>0.999	95.47	4.94	0.028	>0.999
<i>Salsola kali</i> -Maui	207.53	10.58	0.0001	>0.999	92.71	6.23	0.037	>0.999
<i>Salicornia europaea</i>	205.02	14.86	0.0002	0.996	54.55	7.93	NS	>0.999
<i>Sarcocornia fruticosa</i>	190.04	21.67	0.0027	0.978	55.43	9.21	NS	0.959
<i>Bassia hyssopifolia</i>	188.26	17.62	0.0026	0.960	67.32	9.44	NS	0.983
<i>Bassia scoparia</i>	187.80	9.01	0.0036	>0.999	68.09	9.72	NS	0.923
<i>Nitrophila occidentalis</i>	184.10	16.80	0.0070	>0.999	-----	-----	-----	-----
<i>Halothamnus subaphyllus</i>	176.36	18.05	0.0152	>0.999	82.58	8.10	NS	0.995

66 species evaluated

7 susceptible (non-zero) species; all *Salsola* spp.

CV= 4.99 to 36.74%

89 species evaluated

30 susceptible (non-zero) species

CV= 3.53 to 43.32%

# BLUPs and the binary/binomial case

- Data converted to “1” or “0”
- Disease vs no disease
- Successes/attempts =  $x$ , e.g., 0.10, 0.20, 0.90, etc.
- Proportions of 0 and 1 set to 0.01 and 0.99, respectively.
- Logit transformation:  $\log(x/1-x)$  or  $\log(\text{odds})$

# Odds ratio

- Odds ratio= the natural logarithm to the power of the BLUP for each species, i.e.,  $e^{\text{BLUP}}$  for logit values
- A probability of whether disease is likely
- Odds ratios  $> 1$  indicate disease occurrence is likely
- The larger the odds ratio the greater the likelihood of disease

# BLUPs

Genus species <sup>1</sup>	CGS				<i>Uromyces salsolae</i>			
	BLUP (Logit)	Standard error of prediction	Pr> t	Odds ratio	BLUP (Logit)	Standard error of prediction	Pr> t	Odds ratio
<i>Salsola kali</i> -U.K.	12.11	2.51	<0.0001	79.12	8.49	2.51	0.002	9.19
<i>Salsola tragus</i>	12.00	2.51	<0.0001	70.67	8.31	2.51	0.002	7.49
<i>Salsola collina</i>	10.94	2.55	<0.0001	24.48	7.12	2.52	0.007	1.62
<i>Salsola paulsenii</i>	9.94	2.77	0.0005	8.98	7.22	2.70	0.011	2.71
<i>Salsola kali</i> -Akhani	9.85	2.77	0.0006	8.24	7.24	2.70	0.011	2.78
<i>Salicornia bigelovii</i>	10.53	2.68	0.0002	16.31	0.06	2.69	NS	0.01
<i>Salsola australis</i>	8.76	2.55	0.0009	2.78	6.82	2.71	0.016	1.64
<i>Salsola kali</i> -Maui	8.81	2.57	0.0009	2.92	6.46	2.61	0.017	0.78
<i>Salicornia europaea</i>	9.46	2.64	0.0005	5.56	-0.01	2.69	NS	0.01
<i>Sarcocornia fruticosa</i>	7.93	2.92	0.0078	1.21	0.39	2.80	NS	0.02
<i>Bassia hyssopifolia</i>	9.27	2.86	0.0016	4.61	2.72	2.81	NS	0.08
<i>Bassia scoparia</i>	8.99	2.95	0.0030	3.50	2.54	2.88	NS	0.07
<i>Nitrophila occidentalis</i>	8.09	3.08	0.0101	1.42	-----	-----	-----	-----
<i>Halothamnus subaphyllus</i>	6.80	3.18	0.0351	0.39	4.62	3.05	NS	0.39

89 species evaluated

26 species with significant disease incidence

15 species with odds ratios greater than 1 -5 native spp.

66 species evaluated

8 species with significant disease incidence

7 species with odds ratios greater than 1 -no native spp.

# Why?

- The large number of inter-specific relationships place the disease reaction of each species in context, genetically and mathematically, with all species analyzed.
- A spider-web analogy is apt: in the case of CGS there are  $89 \times 89 = 7921$  interconnected nodes in the G matrix, and a force, e.g., disease, applied to any node “pulls” the other nodes, and web, in that direction. Forces at other nodes “pull” in opposing directions.
- Or, in the case of CGS, BLUPs reflect  $59 \times 59$  (species with data)  $\div 2 = 1,740$  fractional replications based on the genetic inter-relationships among these species.
- BLUPs reflect the disease reactions of each species plus the disease reactions of all of the other inter-related species.

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- **BLUPs are safer (higher power) than lsmeans**
- **BLUPs are environmentally independent**

# Advantages continued

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- **BLUPs predict species performance (vs. averages from the material tested in a greenhouse)**
- **The mixed model equations and BLUPs can be used to construct test plant lists**

# Further Implications

- Useable with any agent and any target
- Useable in *ex-post* analyses
- Useable with historical data
- Useable by anyone

# Questions

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# Questions

- Why is the average of a few leaf spots/chews/eggs (or lack thereof) on an infinitesimal sample of a species deemed representative of the species as a whole?
- Why doesn't there seem to be a standard objective criterion for evaluation of susceptibility/damage?
- Why isn't probability of susceptibility/damage that criterion?

# Questions

- Is this species susceptible based on one pustule on one leaf of one plant in a greenhouse test?



- Probably not: BLUP=63.3;  $Pr>|t|=0.95$ ; odds ratio=0.013
- Doubtful that even the plant is susceptible
- Initial APHIS feedback seems to disagree



# Questions

- Whatever happened to the scientific method and rigorous hypothesis testing?

$$H_0: \mu_1 = \mu_2 = \mu_n = 0$$

- Is this not necessary in biological control of weeds?
- Or is science simply not necessary for regulators?



# Recent Publications

- Berner, D.K. 2010. BLUP, a new paradigm in host-range determination. *Biological Control* 53: 143-152.
- Berner, D. K., W. L. Bruckart, C. A. Cavin, J. L. Michael, M. L. Carter, and D. G. Luster. 2009. Best linear unbiased prediction of host range of the facultative parasite *Colletotrichum gloeosporioides* f. sp. *salsolae*, a potential biological control agent of Russian thistle. *Biological Control* 51:158-168.
- Berner, D. K., W. L. Bruckart, C. A. Cavin, and J. L. Michael. 2009. Mixed model analysis combining disease ratings and DNA sequences to determine host range of *Uromyces salsolae* for biological control of Russian thistle. *Biological Control* 49:68-76.

# Using BLUPs to construct non-target test lists

Target →

Genus species	BLUP	S.E.
<i>Rhaponticum repens</i>	70.73	35.51
<i>Arctium minus</i>	-3.66	36.72
<i>Callistephus chinensis</i>	7.68	36.09
<i>Centaurea montana</i>	5.77	35.67
<i>Plectocephalus rothrockii</i>	0.79	36.05
<i>Cichorium intybus</i>	-3.93	36.15
<i>Cirsium pitcheri</i>	0.79	35.77
<i>Crupina vulgaris</i>	12.83	36.65
<i>Erigeron rhizomatus</i>	16.58	35.85
<i>Helianthis eggertii</i>	3.95	36.33
<i>Helianthis schweini</i>	4.34	36.33
<i>Krigia montana</i>	-11.46	36.09
<i>Picnomon acarna</i>	3.64	36.14
<i>Saussurea alpine</i>	4.80	36.74
<i>Serratula coronata</i>	14.06	36.54
<i>Solidago shortii</i>	11.46	36.06
<i>Stokesia laevis</i>	15.41	35.44
<i>Carthamus tinctorius</i>	15.86	36.06
<i>Cynara scolymus</i>	1.55	35.86
<i>Plectocephalus americana</i>	4.25	35.28
<i>Carduus tenuifloris</i>	-6.45	35.75
<i>Carduus thoermeri</i>	-3.97	35.72
<i>Centaurea calcitrapa</i>	-11.42	35.79

= no data  
 = possible susceptible

- Iteratively test, re-analyze, re-test until suspect reactions clarified
- Iteratively include more closely related species and sequences in each test-and-analysis cycle

# Is “what you see” what you get?

- Simple random sampling of test material assumes that the samples are truly representative of the species
- False positives and negatives, from a species perspective, are a real probability
- No indication of probability of “correct” evaluation

# Is “what you see” what you get?

- In the case of CGS, each BLUP in our evaluation is based on 89 species' interactions in addition to observed data
- This greatly increases the probability that the BLUPs are representative of the species
- Probabilities of taxon differences can be tested